In the Specification:

Please amend the substitute specification submitted with the Amendment in Response to the August 15, 2002, Office Action on January 15, 2003, as follows:

Please replace the paragraph at page 1, lines 5-18 with the following:

-- This application is a Divisional application of U.S. Serial No. 09/564,329, filed May 3, 2000, now issued U.S. Patent No. 6,541,212, issued on April 1, 2003, which is a continuation-in-part (CIP) of U.S. Serial No. 09/359,326, filed July 20, 1999, now abandoned, which is a CIP of U.S. Serial No. 09/318,503, filed May 25, 1999, now issued U.S. Patent No. 6,261,791, issued on July 17, 2001, ~~based on U.S. Serial No. 09/318,503, filed May 25, 1999,~~ which is a CIP of U.S. Serial No. 09/251,835, filed February 17, 1999, now issued U.S. Patent No. 6,261,789, issued on July 17, 2001, ~~based on U.S. Serial No. 09/251,835, filed February 17, 1999,~~ which is a CIP of U.S. Serial No. 09/203,939, filed December 2, 1998, now issued U.S. Patent No. 6,258,939, issued on July 10, 2001, ~~based on U.S. Serial No. 09/203,939, filed December 2, 1998,~~ which is a CIP of U.S. Serial No. 09/038,261, filed March 10, 1998, now issued U.S. Patent No. 6,267,960, issued on July 31, 2001, ~~based on U.S. Serial No. 09/038,261, filed March 10, 1998~~; claiming the priority of provisional applications, U.S. Serial No. 60/228,816, filed March 10, 1997; U. S. Serial No. 60/071,141, filed January 12, 1998 and; U. S. Serial No. 60/074,675, filed February 13, 1998. This application which is a divisional of U.S. Serial No. 09/564,329, filed May 3, 2000, now issued U.S. Patent No. 6,541,212, issued on April 1, 2003, further claims the benefit of the filing dates of U.S. Serial Nos. 60/124,658, filed March 16, 1999; 60/120,536 filed February 17, 1999; and 60/113,230 filed December 21, 1998. The contents of all of the foregoing applications are incorporated by reference into the present application. --

Please replace the paragraph at page 41, lines 22-25 with the following:

--Fragments of human *PSCA* that are particularly useful as selective hybridization probes or PCR primers can be readily identified from the entire *PSCA* sequence using art-known methods. One set of PCR primers that are useful for RT-PCR analysis comprise 5' -

TGCTTGCCCTGTTGATGGCAG (SEQ ID NO. 19) - and 3' - CCAGAGCAGCAGGCCGAGTGCA - (SEQ ID NO. 20). --

Please replace the paragraph at page 70, lines 3-9 with the following:

-- In preferred embodiments, DNA fragments of 9kb, 6kb, 3kb, and 1kb derived from the 5' upstream region of the PSCA gene, as shown in Figure 42, were produced by techniques described herein. The 9kb PSCA upstream region (pEGFP—PSCA) is involved with gene regulatory activity and was deposited on May 17, 1999 with the American Type Culture Collection (ATCC), 12301 Parklawn Drive, Rockville, MD 20852 and has there been identified as follows ATCC No. PTA-80. The 9 kb fragment was obtained by amplification using a T7 primer and RIhPSCA3'-5 (5'-gggaattcgcacagccttcagggtc-3') (SEQ ID NO. 21). --

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Please replace at page 92, line 27 with the following:

-- 5'- GGAGAATTCATGGCACTGCCCTGCTGTGCTAC (SEQ ID NO. 22).--

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Please replace at page 93, line 28 with the following:

-- 3'-GGAGAATTCCTAATGGGCCCCGCTGGCGTT (SEQ ID NO. 23). --

Please replace the paragraph at page 110, lines 15-32 continuing to page 111, lines 1-7 with the following:

-- The reporter gene vectors are depicted in Figure 42 and were constructed as follows. The 14 kb Not I fragment was sub-cloned from the λ vector into a Bluescript KS vector (Stratagene), resulting in the pBSKS-PSCA (14kb) construct. The PSCA upstream sequence was subcloned from pBSKS-PSCA (14 kb) by PCR amplification using a primer corresponding to the T7 sequence contained within the Bluescript vector, and a primer corresponding to a sequence contained within PSCA exon 1 (primer H3hPSCA3'-5, the sequence of this primer is as follows: The sequence of H3hPSCA3'-5 is 5'-gggaagcttgcacagccttcagggtc-3' (SEQ ID NO 24). The primer corresponding to PSCA exon 1 contained an introduced HindIII sequence to allow further subcloning following

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PCR amplification. The resulting amplified fragment was digested with HindIII and was subcloned into the pGL3-basic vector (Promega) to generate pGL3-PSCA (7 kb) which was used to generate a series of deletion reporter gene constructs containing varying lengths of PSCA upstream sequences operatively linked to the luciferase gene (Figure 42). The deleted portions of the PSCA upstream regions were obtained by subcloning restriction fragments from pGL3-PSCA (7 kb). The PSCA upstream region between –9 kb and –7 kb was subcloned from the pBSKS-PSCA (14 kb) construct, the Not I site was converted into a blunt end by Klenow and the fragment was cloned into the SacI/HindIII sites of pGL-PSCA (7 kb) in order to obtain the pGL3-PSCA (9 kb) construct. The reference to the sequences upstream of the PSCA coding region, such as –9 kb and –6 kb (etc.), are relative to the ATG start translation codon. The reporter gene constructs pGL3-PSCA (9 kb), pGL3-PSCA (6 kb), pGL3-PSCA (3 kb), and pGL3-PSCA (1 kb) were operatively linked to the luciferase gene (Figure 42). Plasmid, pGL3-CMV contains the cytomegalovirus promoter (Boshart, M. et al., 1985 *Cell* 41:521-530) linked to the luciferase gene and was used as a positive control. Also, plasmid pGL3 contains no promoter sequence and was used as a negative control plasmid. --

Please replace at page 131, line 25 with the following:

-- HLEAD.1: ggc gat atc cac cat ggR atg Sag ctg Kgt Mat Sct ctt (SEQ ID NO. 25). --

Please replace at page 131, line 27 with the following:

-- CH3': agg gaa ttc aYc tcc aca cac agg RRc cag tgg ata gac (SEQ ID NO. 26). --

Please replace at page 131, line 31 with the following:

-- HLEAD.2: ggg gat atc cac cat gRa ctt cgg gYt gag ctK ggt ttt (SEQ ID NO. 27). --